

Application News

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Liquid Chromatography Mass Spectrometry

Multi-Residue Analysis of 18 Regulated Mycotoxins by LC/MS/MS

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Mycotoxins are one of the most important contaminants in food and feed due to their widespread distribution in the environment and toxic effects on humans and animals.¹⁾ Structurally, mycotoxins are a very diverse group with a wide range of physicochemical properties and low molecular weights.²⁾ They are produced by fungi (mould) frequently found on agricultural produce, and are often not visible to the naked eye.³⁾ Some of the most commonly contaminated food stuffs include wheat, oats, rye, corn, barley, rice, nuts and milk.⁴⁾

Due to the risks posed by mycotoxins in food they are regulated globally, including, the EU, US, China, Singapore and Brazil.⁵⁾ In the EU, reporting limits are harmonised in Regulation (EC) No 1886/2006 (amended by (EC) No 1126/2007) and sampling and analysis in Regulation (EC) No 401/2006.

LC/MS/MS is the technique most commonly employed for mycotoxin quantitation in order to achieve the necessary low reporting limits in complex food and feed matrices.

Experimental

Solvent extracts were provided by Scientific Analysis Laboratories (SAL, UK) following validated extraction protocols. Samples were analysed using the Nexera UHPLC and the LCMS-8060 triple quadrupole detector (Table 1). Calibration was performed using ¹³C internal standards spiked during sample extraction. All MRM transitions and associated internal standards for each compound are listed in Table 2. All solvents used during analysis were LCMS quality from Sigma-Aldrich.

Due to the wide range of physical and chemical properties of mycotoxins, different LC/MS/MS methods are typically developed for small groups of compounds with similar properties.

In this application paper a single LC/MS/MS method has been developed for the determination of 18 mycotoxins in food safety. Limits of quantification were at or below the maximum levels set in the EC/1886/2006 document. The scope of the method included Aflatoxins (B1, B2, G1, G2), Fumonisin (B1, B2, B3), Ochratoxin A (OTA) and Trichothecenes (3-acetyldeoxynivalenol (3AcDON), 15-acetyldeoxynivalenol (15AcDON), Deoxynivalenol (DON), Diacetoxyscripanol (DAS), Fusarenon-X (FUS X), HT-2, Neosolaninol (NEO), Nivalenol (NIV), T2, Zearalenone (ZON)) with an analysis cycle time of 12.5 minutes.

Table 1 Analytical Conditions

| | |
|--------------------|--|
| UHPLC | : Nexera LC System |
| Mobile Phase | : A; Water with additives B; Methanol with additives |
| Column | : Reversed phase column (100 mm L x 2.1 mm I.D.) |
| Column Temperature | : 40 °C |
| Flowrate | : 0.4 mL/minute |
| Gradient | : B. Conc 15 % (0 min) → 25 % (1 min) → 40 % (2 min) → 41 % (4.5 min) → 100 % (7.5 - 10.0 min) → 15 % (10.10 min) → Stop (12.5 min) |
| LC-MS/MS | : LCMS-8060 |
| Dwell Time | : 10 to 40 msec. |
| Pause Time | : 1 msec. |
| Ionisation Mode | : ESI +/- |
| Polarity Switching | : 5 msec. |

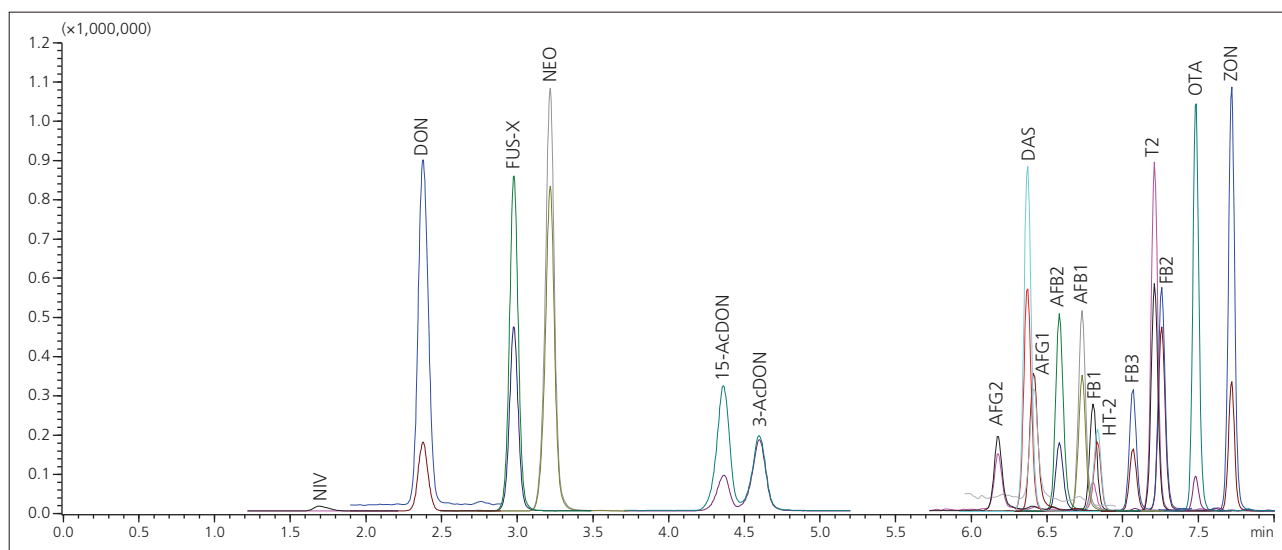


Fig. 1 MRM Chromatograms of 18 Mycotoxins

AFB1 (aflatoxin B1; 1 µg/kg), AFB2 (aflatoxin B2; 1 µg/kg), AFG1 (aflatoxin G1; 1 µg/kg), AFG2 (aflatoxin G2; 1 µg/kg), OTA (ochratoxin A; 4 µg/kg), FB1 (fumonisin B1; 100 µg/kg), FB2 (fumonisin B2; 100 µg/kg), FB3 (fumonisin B3; 100 µg/kg), 15-AcDON (15-acetyldeoxynivalenol; 100 µg/kg), 3-AcDON (3-acetyldeoxynivalenol; 100 µg/kg), DON (deoxynivalenol; 100 µg/kg), DAS (diacetoxyscripanol; 100 µg/kg), FUS-X (fusarenon-X; 100 µg/kg), HT-2 (100 µg/kg), T-2 (100 µg/kg), NEO (neosolaninol; 100 µg/kg), NIV (nivalenol; 100 µg/kg), ZON (zearalenone; 100 µg/kg).

For clarity only 2 MRM transitions are displayed per compound and the following MRM chromatograms were changed; neosolaninol (x0.3), T2 (x0.3), aflatoxins (x3), fumonisins (x2).

Table 2 All MRM's Measured in the Mycotoxin Method and Corresponding Calibration Range and R² Result

| Compound name | Parent ion | Ret. Time (mins) | MRM 1 | MRM 2 | MRM 3 | ISTD | Calibration range µg/kg | R ² |
|---------------------------------|--------------------------------------|------------------|-----------|-----------|-----------|--------------------------------|-------------------------|----------------|
| 1 Aflatoxin B1 | [M+H] ⁺ | 6.773 | 313 > 241 | 313 > 285 | 313 > 269 | ¹³ C Aflatoxin B1 | 0.1 - 10 | 0.9988 |
| 2 Aflatoxin B2 | [M+H] ⁺ | 6.621 | 315 > 259 | 315 > 287 | 315 > 243 | ¹³ C Aflatoxin B2 | 0.1 - 10 | 0.9995 |
| 3 Aflatoxin G1 | [M+H] ⁺ | 6.453 | 329 > 243 | 329 > 200 | | ¹³ C Aflatoxin G1 | 0.1 - 10 | 0.9998 |
| 4 Aflatoxin G2 | [M+H] ⁺ | 6.219 | 331 > 245 | 331 > 285 | | ¹³ C Aflatoxin G2 | 0.1 - 10 | 0.9965 |
| 5 Ochratoxin A | [M+H] ⁺ | 7.509 | 404 > 239 | 404 > 221 | 404 > 358 | ¹³ C Ochratoxin A | 0.4 - 40 | 0.9969 |
| 6 Fumonisin B1 | [M+H] ⁺ | 6.811 | 722 > 352 | 722 > 334 | 722 > 704 | ¹³ C Aflatoxin B2 | 10 - 1000 | 0.9937 |
| 7 Fumonisin B2 | [M+H] ⁺ | 7.260 | 706 > 318 | 706 > 354 | 706 > 688 | ¹³ C Aflatoxin B2 | 10 - 1000 | 0.9998 |
| 8 Fumonisin B3 | [M+H] ⁺ | 7.073 | 706 > 318 | 706 > 354 | 706 > 688 | ¹³ C Aflatoxin B2 | 10 - 1000 | 0.9991 |
| 9 Deoxynivalenol | [M+H] ⁺ | 2.372 | 297 > 279 | 297 > 249 | | ¹³ C Deoxynivalenol | 10 - 1000 | 0.9992 |
| 10 Diacetoxyscirpenol | [M+NH ₄] ⁺ | 6.349 | 384 > 229 | 384 > 307 | 384 > 247 | ¹³ C T2 Toxin | 10 - 1000 | 0.9994 |
| 11 T2 | [M+NH ₄] ⁺ | 7.206 | 484 > 185 | 484 > 215 | 484 > 245 | ¹³ C T2 Toxin | 10 - 1000 | 0.9989 |
| 12 HT-2 | [M+Na] ⁺ | 6.822 | 447 > 345 | 447 > 285 | | ¹³ C T2 Toxin | 10 - 1000 | 1.0000 |
| 13 Nivalenol | [M-CH ₃ COO] ⁻ | 1.684 | 371 > 281 | 371 > 311 | | ¹³ C HT-2 | 10 - 1000 | 0.9991 |
| 14 Neosolaniol | [M+NH ₄] ⁺ | 3.227 | 400 > 215 | 400 > 305 | 400 > 185 | ¹³ C Deoxynivalenol | 10 - 1000 | 0.9995 |
| 15 Fusarenon X | [M+H] ⁺ | 2.986 | 355 > 247 | 355 > 277 | | ¹³ C Deoxynivalenol | 10 - 1000 | 0.9987 |
| 16 Zearalenone | [M-H] ⁻ | 7.711 | 317 > 175 | 317 > 131 | 317 > 273 | ¹³ C T2 Toxin | 10 - 1000 | 0.9985 |
| 17 15-Acetyldeoxynivalenol | [M+H] ⁺ | 4.406 | 339 > 261 | 339 > 297 | | ¹³ C Deoxynivalenol | 10 - 1000 | 1.0000 |
| 18 3-Acetyldeoxynivalenol | [M+H] ⁺ | 4.618 | 339 > 261 | 339 > 297 | | ¹³ C Deoxynivalenol | 10 - 1000 | 0.9986 |
| 19 ¹³ C HT-2 | [M+NH ₄] ⁺ | 6.844 | 464 > 278 | | | | | |
| 20 ¹³ C T2 | [M+NH ₄] ⁺ | 7.228 | 508 > 322 | | | | | |
| 21 ¹³ C Aflatoxin B1 | [M+H] ⁺ | 6.754 | 330 > 301 | | | | | |
| 22 ¹³ C Aflatoxin B2 | [M+H] ⁺ | 6.614 | 332 > 303 | | | | | |
| 23 ¹³ C Aflatoxin G1 | [M+H] ⁺ | 6.435 | 346 > 212 | | | | | |
| 24 ¹³ C Aflatoxin G2 | [M+H] ⁺ | 6.219 | 348 > 259 | | | | | |
| 25 ¹³ C Ochratoxin A | [M+H] ⁺ | 7.516 | 424 > 250 | | | | | |

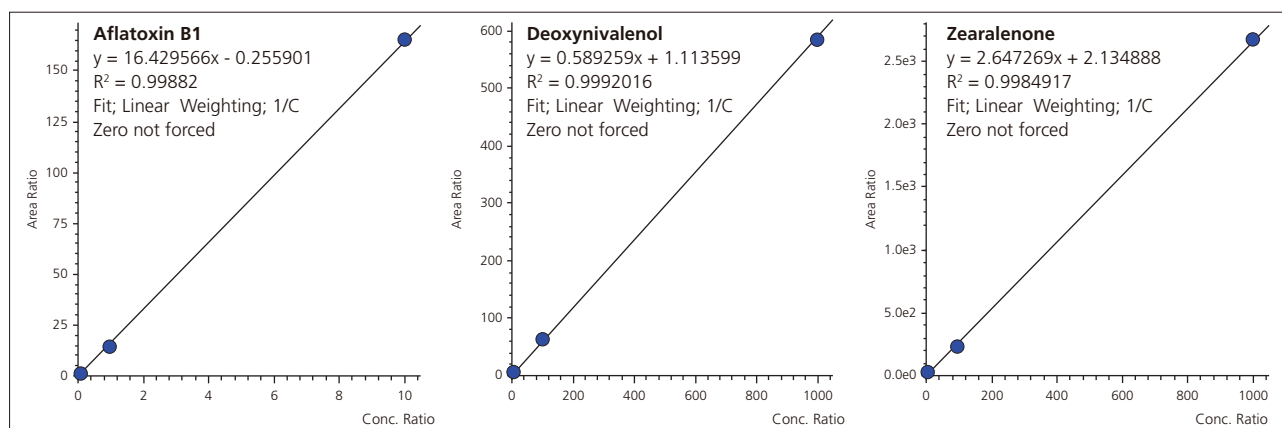


Fig. 2 Calibration Curves for Selected Compounds
Calibration Curves for Aflatoxin (0.1 – 10 µg/kg), Deoxynivalenol (10 – 1000 µg/kg), and Zearalenone (10 – 1000 µg/kg).

Conclusions

In this study a single method has been developed for the analysis of 18 regulated mycotoxins with an injection to injection cycle time of 12.5 minutes. This method achieves the required EU reporting limits (between 0.1 -10 µg/kg) with linear regression

coefficients R² typically greater than 0.998 (Fig. 2 and Table 1). The LC mobile phase, column and gradient were all optimised and provided chromatographic resolution of 15-acetyldeoxynivalenol and 3-acetyldeoxynivalenol.

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